

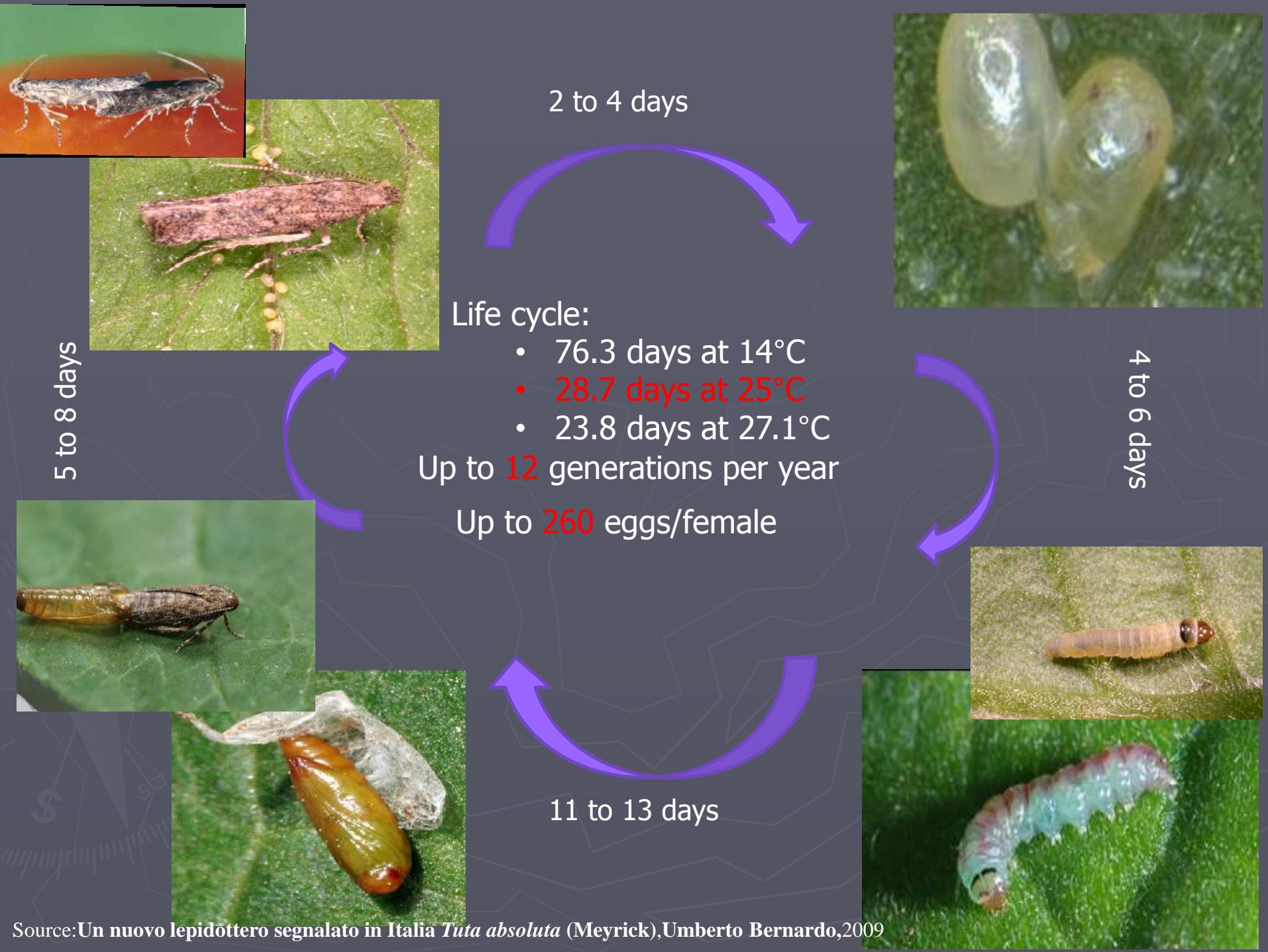


“*Tuta absoluta* (Meyrick) and insecticide resistance ”

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ROTHAMSTED
RESEARCH





Source: Un nuovo lepidottero segnalato in Italia *Tuta absoluta* (Meyricke)

- Reported presence
- Unreported presence
- Present in glasshouses only
- △ Potential *Tuta absoluta* outbreak



Control:

- Organophosphates and pyrethroids were used during the 1970's and 1980's
- In the 1990's, new products introduced such as abamectin, spinosad, tebufonzide, and chlorfenpyr. (Lietti *et al.*, 2005)
- up to 36 insecticide applications per single tomato crop cycle (Picanço *et al.*, 1995)

Registered Insecticides on Tomato used to Control *Tuta absoluta*

MOA Group	Chemical subgroup	Common Name	
Acetylcholinesterase (AChE) inhibitors	1B: Organophosphates	Chlorpyrifos Methamidophos	
Sodium channel modulators	3A: Pyrethroids	Deltamethrin	
Nicotinic acetylcholine receptor (nAChR) agonists	4A: Neonicotinoids	Imidacloprid Thiacloprid	
Nicotinic acetylcholine receptor (nAChR) allosteric activators	5:Spinosyns	Spinosad	
Chloride channel activators	6:Avermectins	Abamectin, emamectin	
Microbial disruptors of insect midgut membranes	11: <i>Bacillus thuringiensis</i>	<i>Bacillus thuringiensis</i>	
Uncouplers of oxidative phosphorylation via disruption of the proton gradient	13:Chlorfenapyr	Chlorfenapyr	
Inhibitors of chitin biosynthesis, type 0	15:Benzoylureas	Iufenuron	
Voltage-dependent sodium channel blockers	22A : indoxacarb 22B: Metaflumizone	Indoxacarb Metaflumizone	
Ryanodine receptor modulators	28: Diamides	Chlorantraniliprole Flubendiamide	
Unknown or act on multiple targets	Azadirachtin	Azadirachtin	

Control failures :

- Brazil : abamectin, cartap, methamidophos and permethrin (Siqueira et al., 2000)
- Chile: deltamethrin, metamidophos, esfenvalerate, lambda cyhalothrin and mevinphos (Salazar and Araya ,1997)
- Argentine: abamectin and Deltamethrin (tolerance reported by Lietti et al., 2005)
- ▶ Gerson et al., 2011 reported evidence of control failures for bifenthrin, permethrin, diflubenzuron, teflubenzuron, triflumuron and *B.thuringiensis*, moderate levels of resistance to indoxacarb and low resistance levels for bifenthrin and permethrin, abamectin, spinosad, *B. thuringiensis* and the mixture of deltamethrin and triazophos.

Questions

“What is the actual status of insecticide resistance of *tuta absoluta* in the Mediterranean region and what kind of Resistance Management Model should be implemented to achieve a sustainable control of the insect ?”

What are the levels of resistance of *tuta absoluta* to the existing and used pesticides in the Mediterranean basin?

What are the risks for the appearance of new insecticide resistance?

What is the most suitable Resistance Management Model to be adopted for the local context?

How can the model be successfully implemented in different cropping systems?

► Bio-assays: leaf dip

A set of leaf dip bioassays has been carried out to evaluate the susceptibility of the reared strains of *Tuta absoluta*.

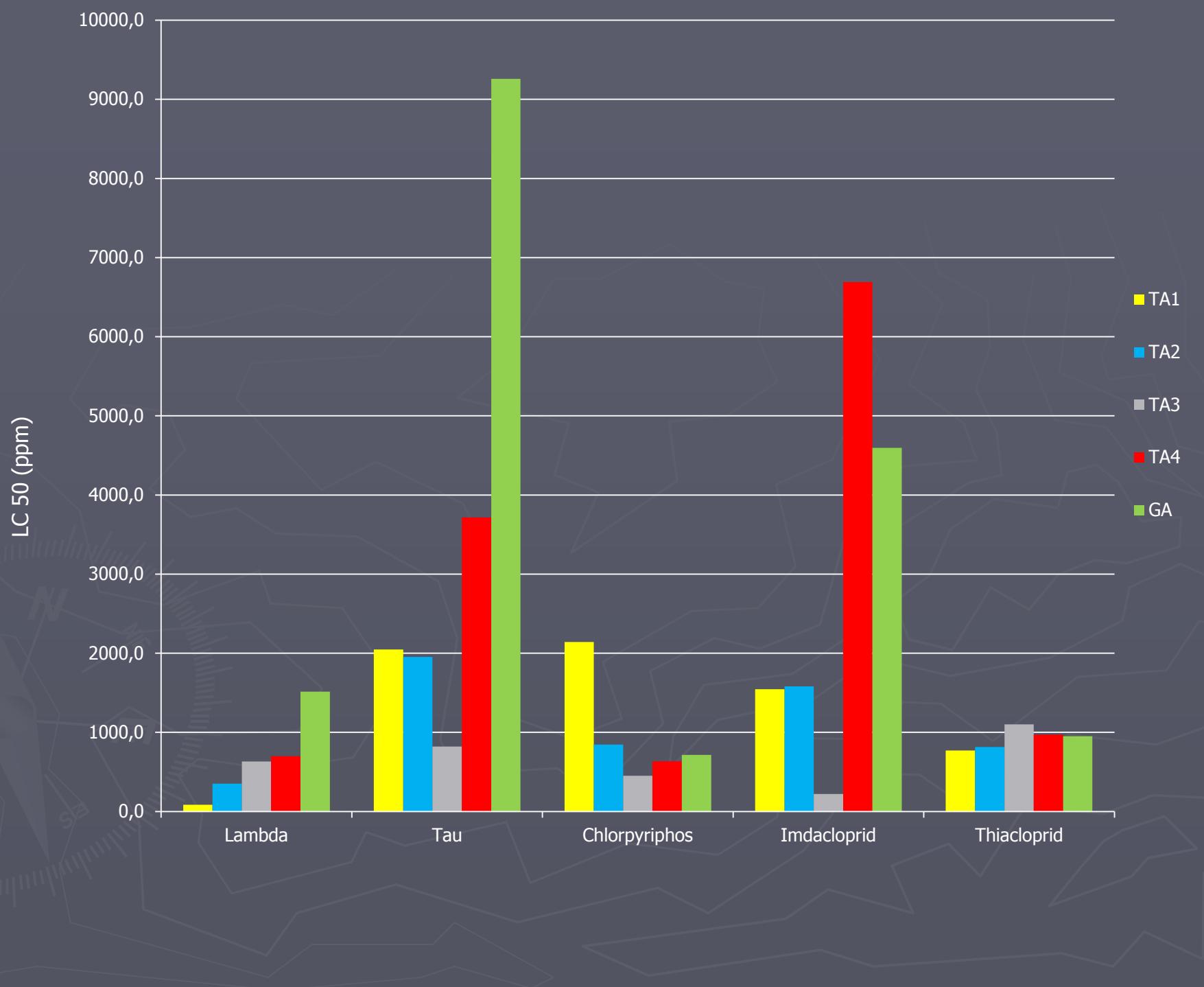
- ▶ The tomato leaves were immersed in six different concentrations insecticide and allowed to air dry for at least one hour.
- ▶ One to two leaves were placed on slightly moistened filter paper inside Petri dishes (90mm diameter × 20 mm height).
- ▶ Fifteen to twenty L2 instars were placed on the leaves in each Petri dish then placed in controlled conditions of temperature (26 ± 2 °C) and light (16h/8h).
- ▶ Three replicates were used for each concentration of insecticide. Each bioassay was scored after 48 hours.
- ▶ The larvae is counted as dead if it shows signs of uncoordinated movements, cannot flip back to the crawling position or cannot crawl for a distance at least equal to the double of its length



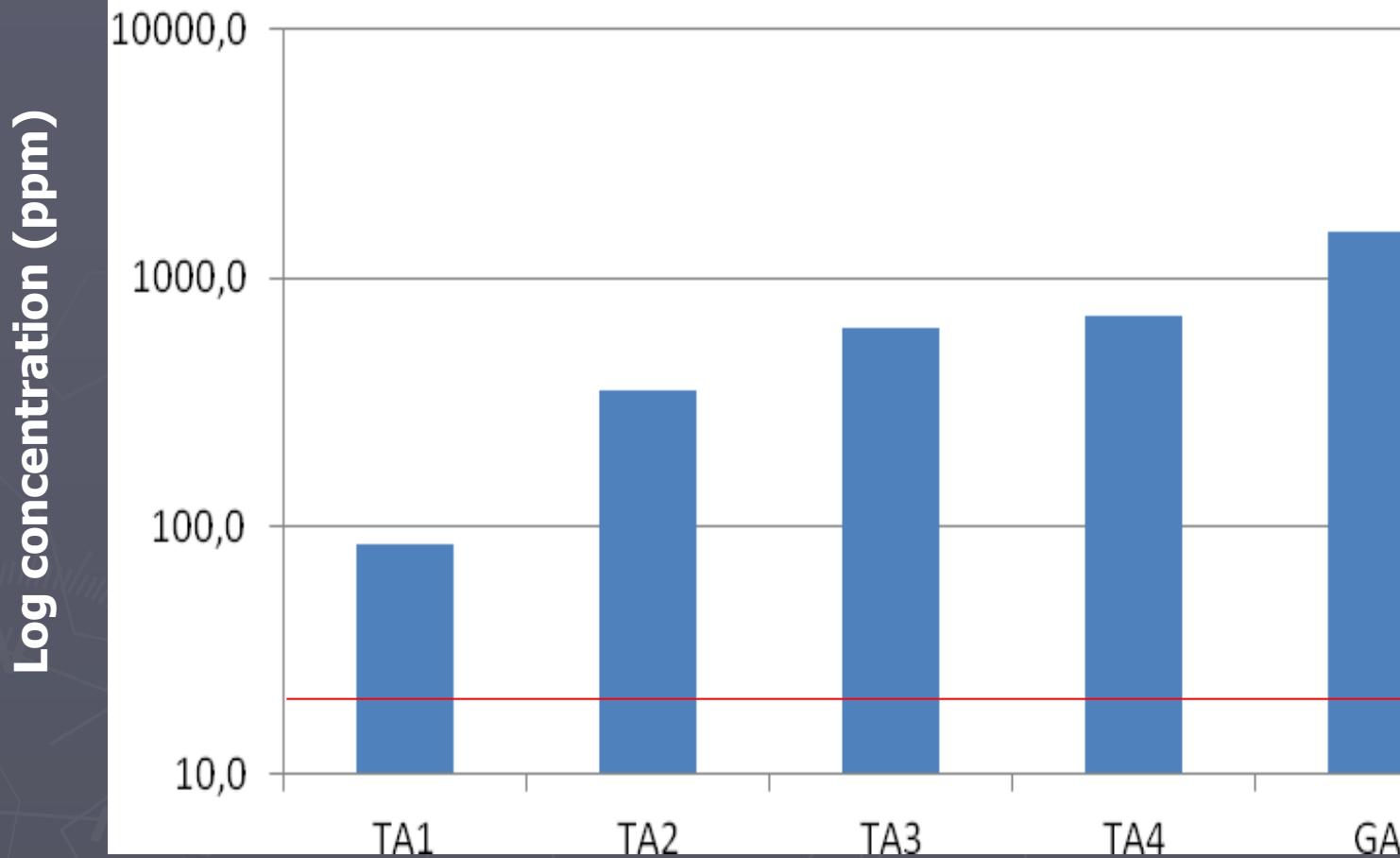
- ▶ *Tuta absoluta* strains:
 - TA1, TA2, TA3, TA4, GA
 - No susceptible strain!!!
 - Highest recommended field rates
- ▶ Chemicals:



Class	Compound
Pyrethroids	Lambda Cyhalothrin
Pyrethroids	Tau fluvalinate
Organophosphates	Chlorpyrifos
Neonicotinoid	Imidacloprid
Neonicotinoid	Thiacloprid



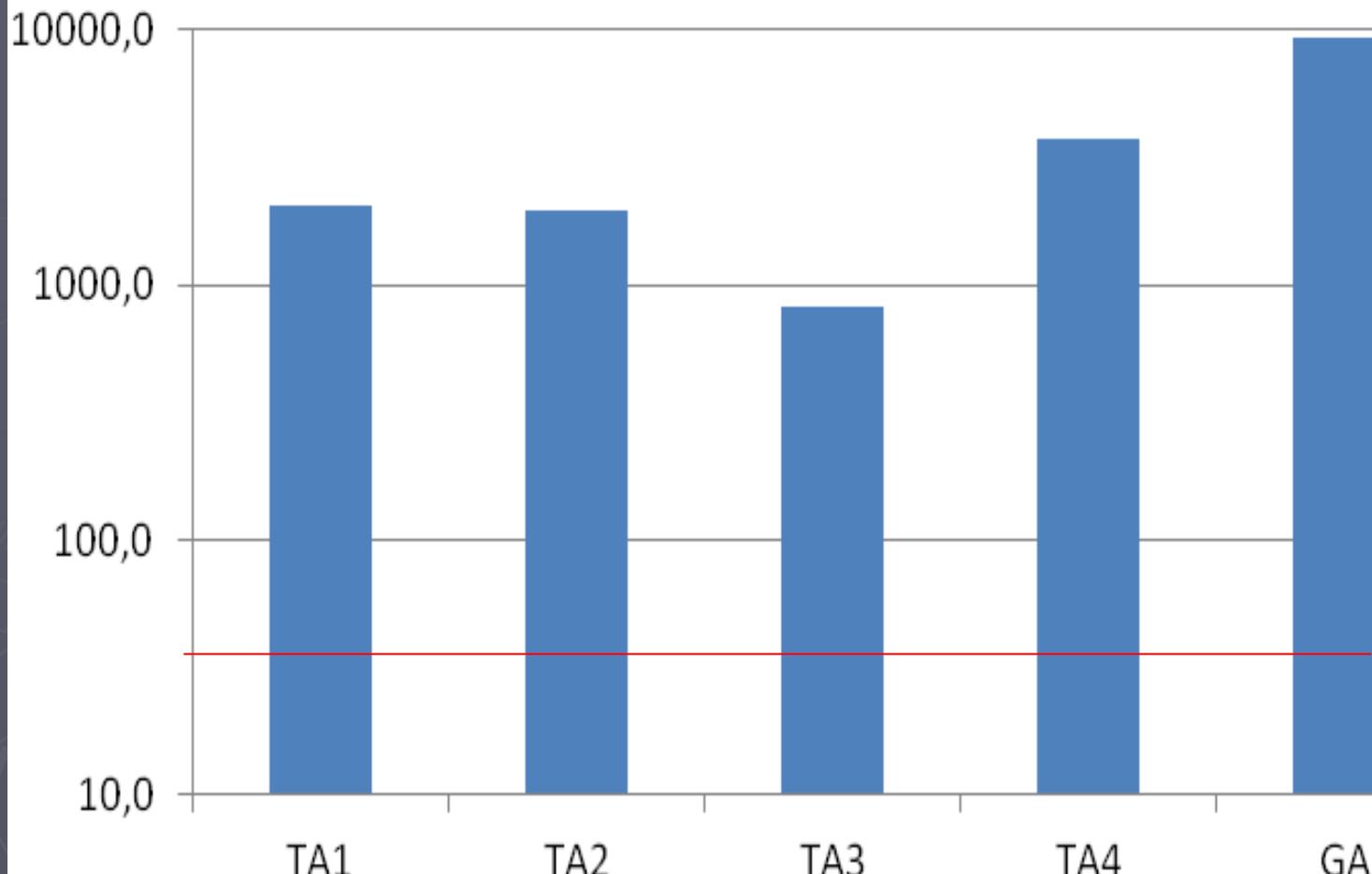
Lambda cyhalothrin



Lambda cyhalothrin
LC50 estimates (Log concentrations)
for five different *T. absoluta* strains.

Tau

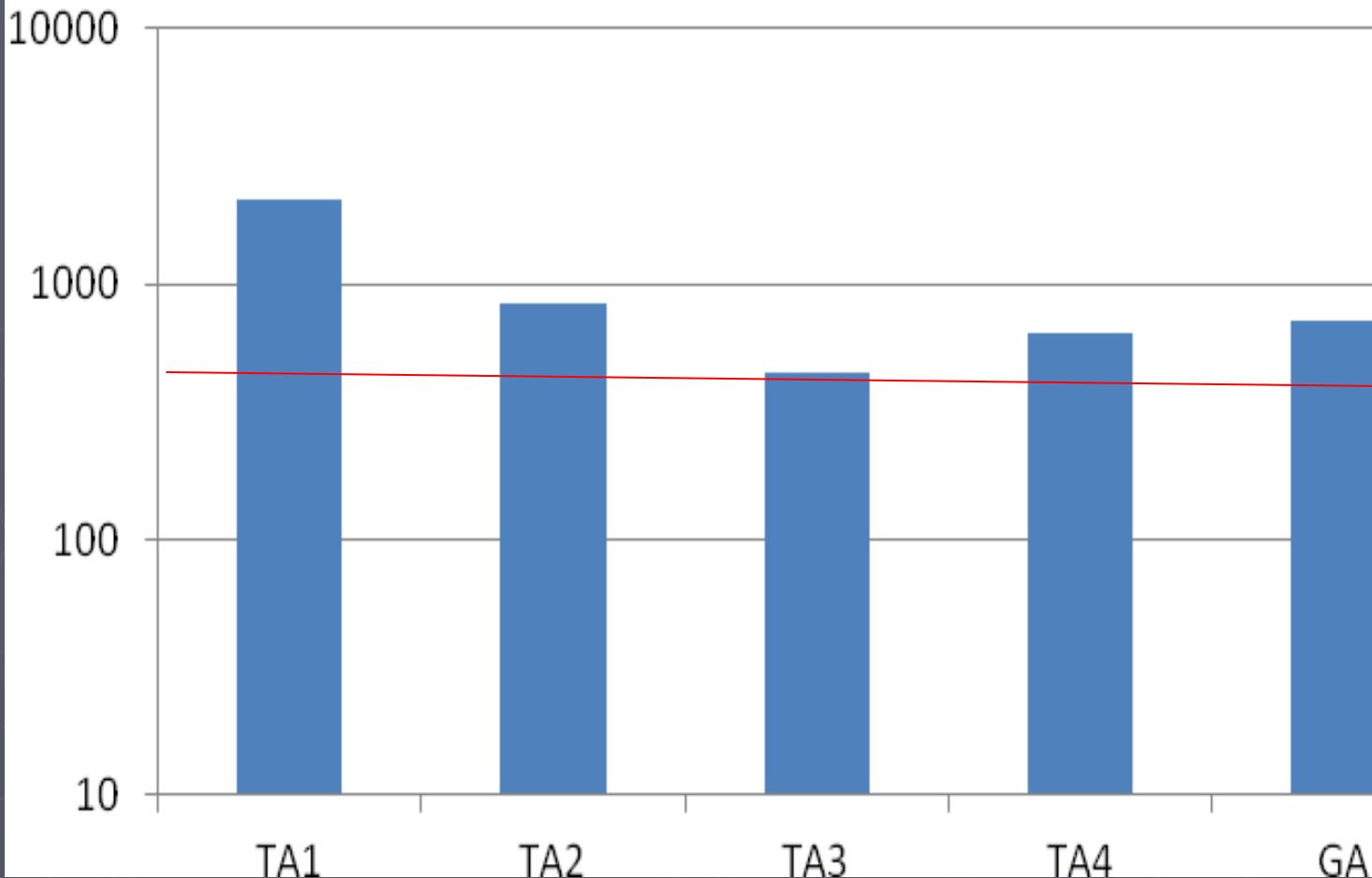
Log concentration (ppm)



**Tau fluvalinate
LC50 estimates (Log concentrations)
for five different *T. absoluta* strains.**

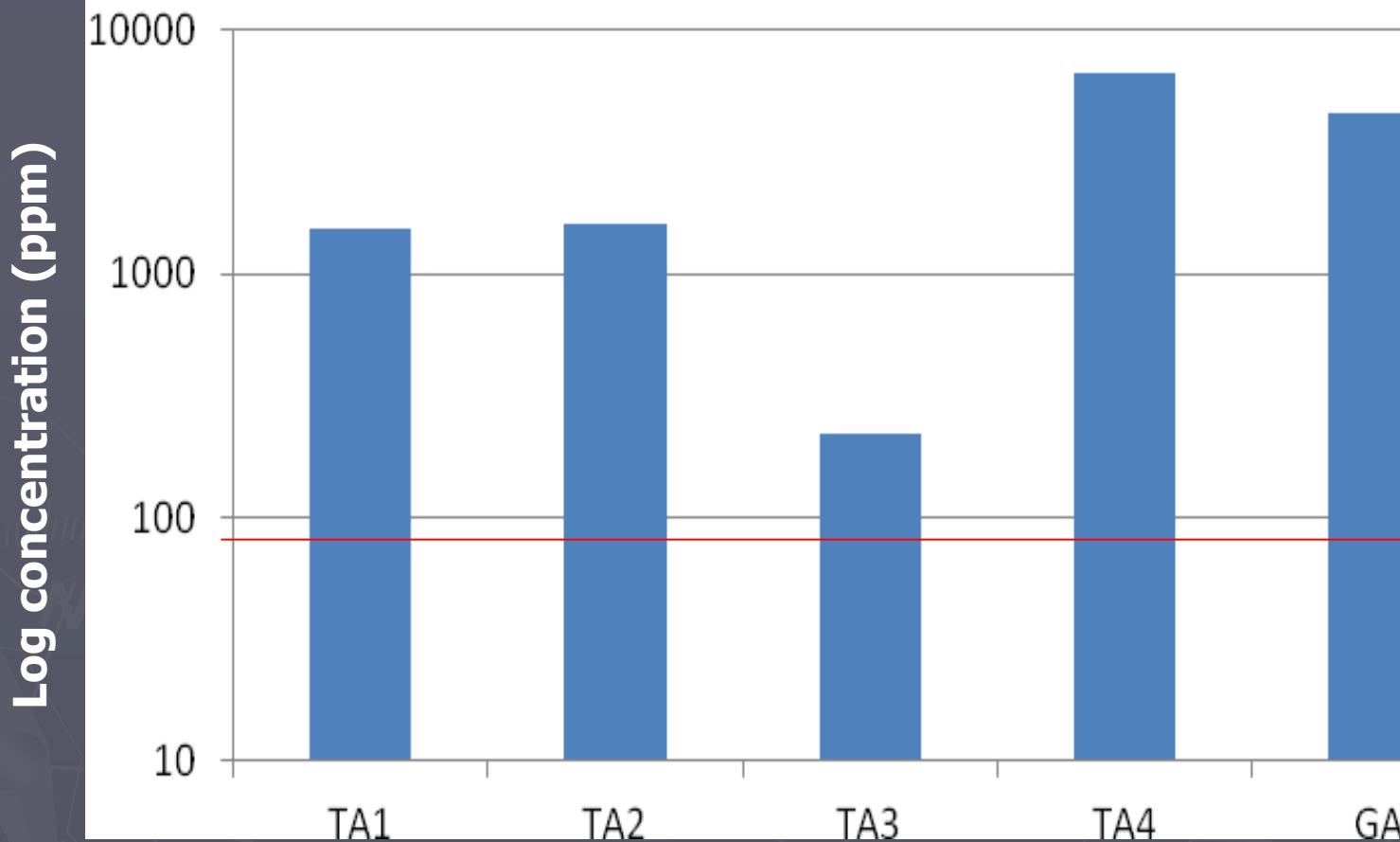
Chlorpyriphos

Log concentration (ppm)



Chlorpyriphos
LC50 estimates (Log concentrations)
for five different *T. absoluta* strains.

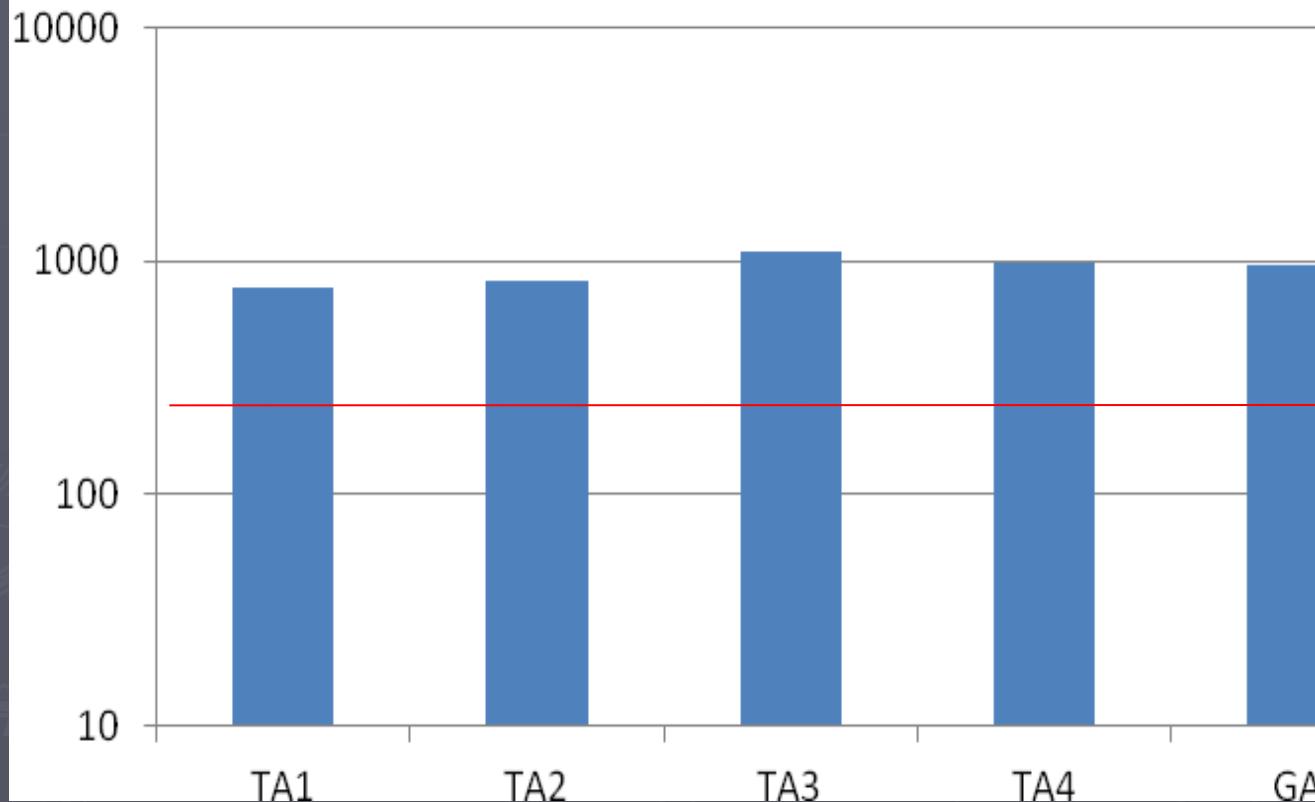
Imidacloprid



Imidacloprid
LC50 estimates (Log concentrations)
for five different *T. absoluta* strains.

Thiacloprid

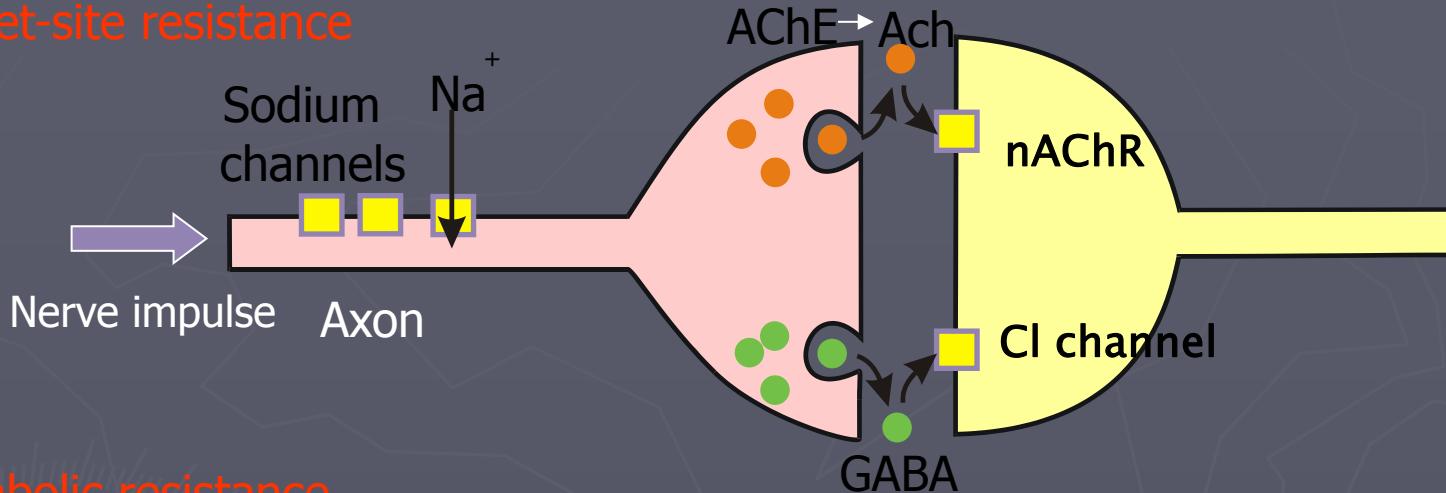
Log concentration (ppm)



Thiacloprid
LC50 estimates (Log concentrations)
for five different *T. absoluta* strains.

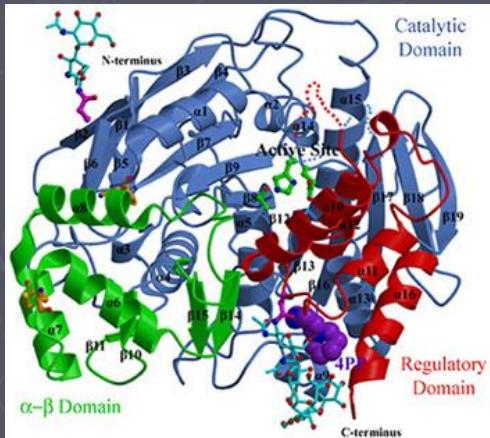
mechanisms of resistance

Target-site resistance

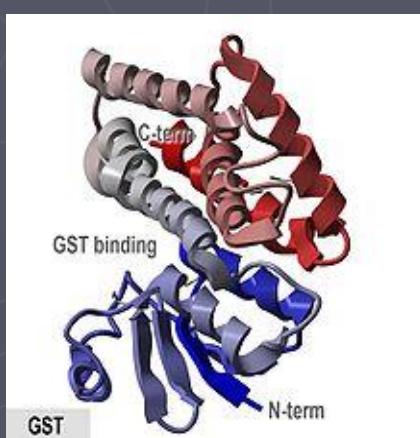


Metabolic resistance

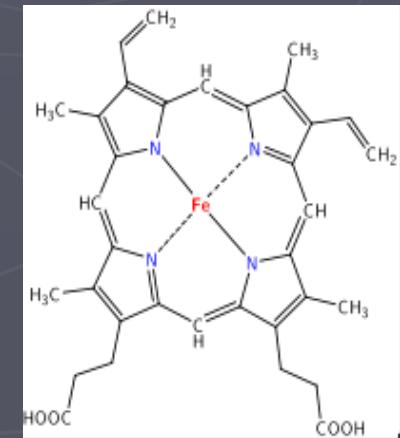
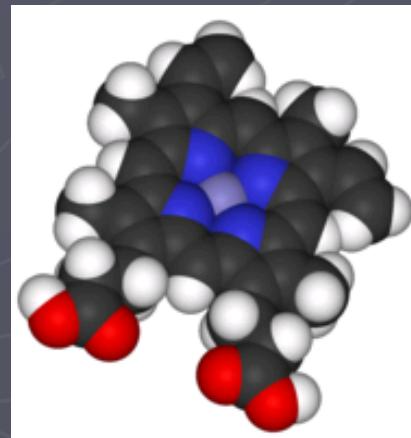
Carboxylesterases



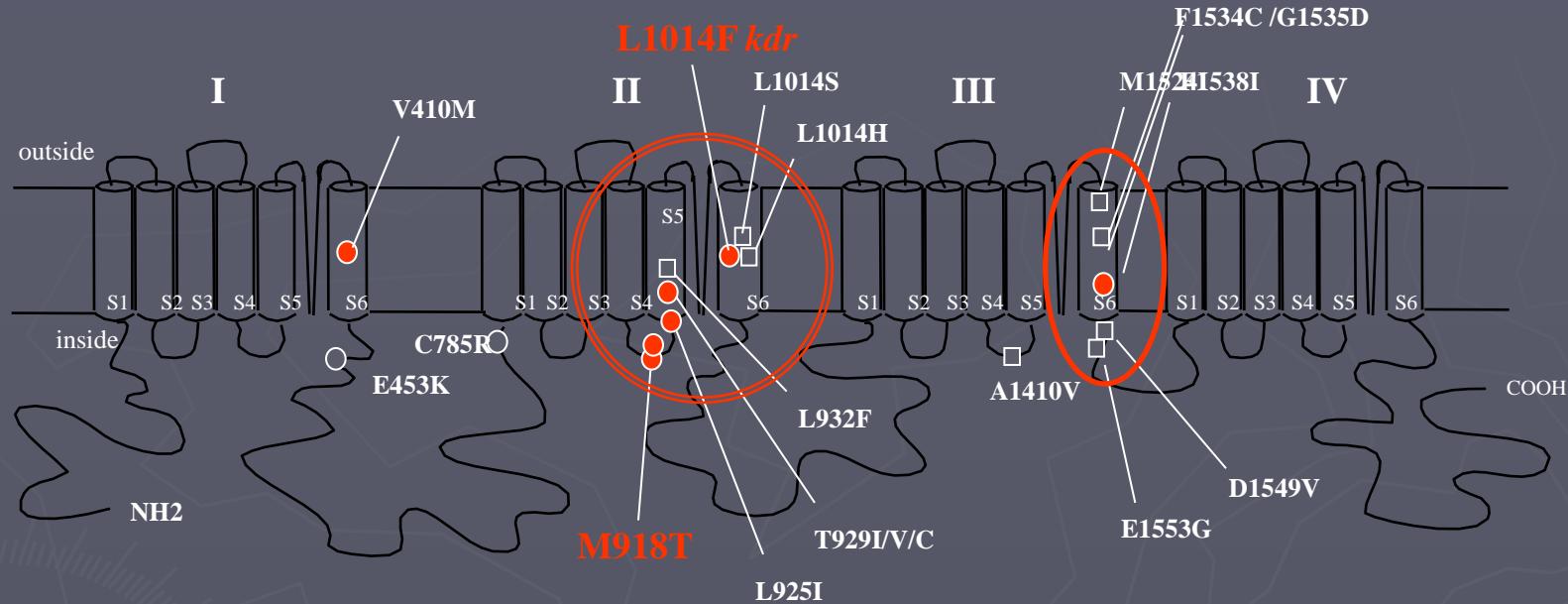
Glutathione S-transferases



Cytochrome P450s

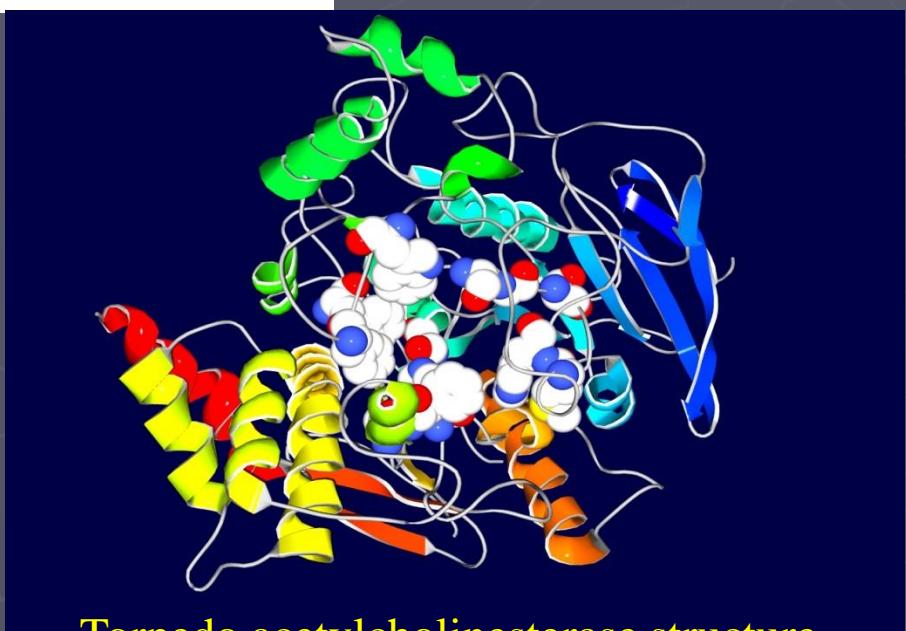
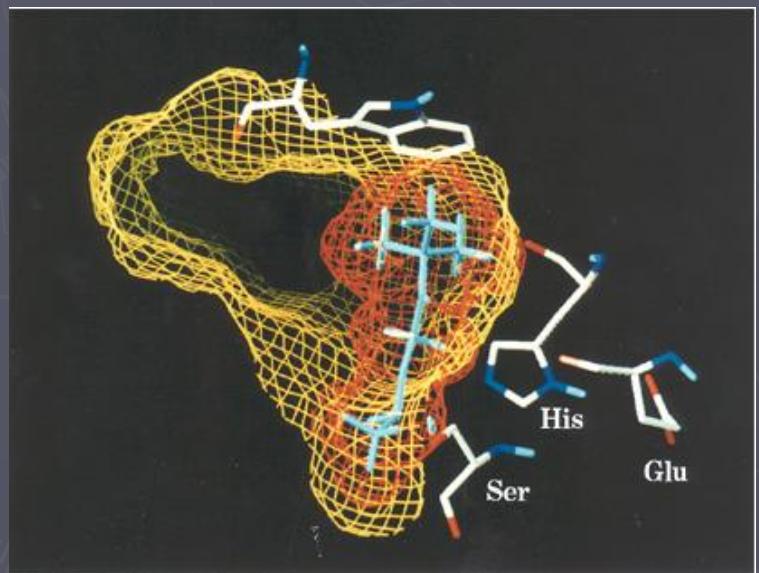
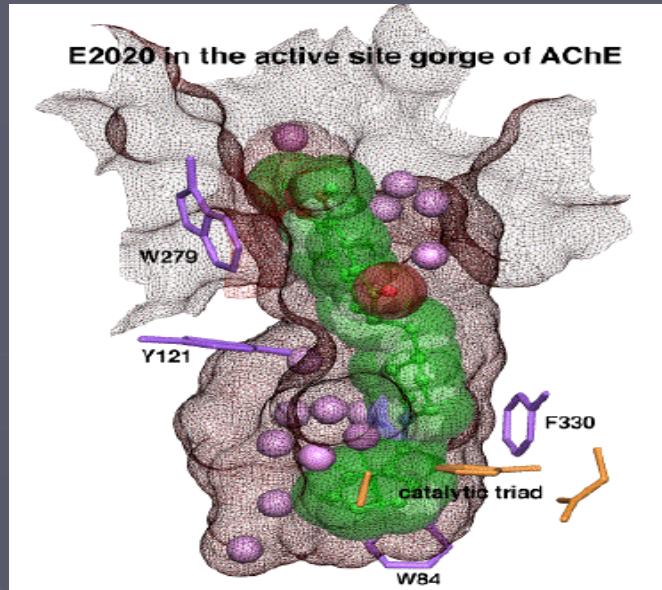


Sodium channel and kdr/super-kdr mutations



Anopheles gambiae
Aphis gossypii
Bemisia tabaci
Blattella germanica
Boophilus microplus
Ctenocephalides felis
Culex pipiens
Drosophila melanogaster
Aedes aegypti
Liriomyza huidobrensis

Helicoverpa armigera
Heliothis virescens
Hematobia irritans
Leptinotarsa decemlineata
Musca domestica
Myzus persicae
Pediculus capitis
Plutella xylostella
Frankliniella occidentalis
Sarcoptes scabiei



Torpedo acetylcholinesterase structure

Mutations	Position	Insect
E to G	73	only in one drosophila population
• F to L • F to S	78	• <i>Aphis gossypii</i> • has been expressed in <i>Drosophila melanogaster</i> , <i>Lucilia cuprina</i> and <i>Aedes aegypti</i> recombinant enzymes
E to K	82	found only once, in a drosophila population
• G to S • G to S	119	• <i>Culex pipiens</i> , <i>Anopheles gambiae</i> and <i>Anopheles albimanus</i> • <i>Tetranychus urticae</i>
D to E	128	<i>Tetranychus urticae</i>
I to V	129	<i>Drosophila</i> , <i>Bractocera oleae</i> , expressed in <i>Lucilia cuprina</i> and drosophila recombinant enzymes
V to L	151	House fly
A to S	201	<i>Aphis gossypii</i> , <i>Chilo suppressalis</i> , <i>Plutella xylostella</i> , <i>Tetranychus urticae</i> , <i>Bactrocera dorsalis</i>
G to A G to V	227	• Drosophila, • <i>Musca domestica</i> , has been expressed in <i>Drosophila melanogaster</i> , <i>Lucilia cuprina</i> , <i>Aedes aegypti</i> and <i>Musca domestica</i> recombinant enzymes
S to G	238	<i>Leptinotarsa decemlineata</i> ,
F to Y	290	<i>Drosophila</i> , <i>Musca domestica</i> , has been expressed in <i>Drosophila</i> , house fly, <i>Lucilia cuprina</i> and <i>Aedes aegypti</i> recombinant enzymes
G to A	328	House fly and drosophila
S to F S to W S to C	331	• <i>Aphis gossypii</i> , • <i>Culex tritaeniorhynchus</i> • <i>Tetranychus urticae</i>
G to S	396	<i>Bactrocera oleae</i>
		(From Fournier, 2005 with amendments)

Tuta absoluta	SWPTFNLLISIMGRTMGALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Plutella	SWPTLNLLISIMGRTMGALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Heliotis	SWPTLNLLISIMGRTMGALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Droso	SWPTLNLLISIMGRTMGALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Musca	SWPTLNLLISIMGRTMGALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Myzus	SWPTLNLLISIMGRT <ins>I</ins> GALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Blattella	SWPTLNLLISIMGRTVGALGNLT <ins>FVLCIIIFIFAVMGMQLFIFAVMGMQL</ins>
Tutaabsoluta	FGKNYVDNVDRFPDGDLPRWNFTDFMHSFMIVFRVLCGEWIESMWDCML
Plutella	FGKNYVDHVDRFPDGDLPRWNFTDFMHSFMIVFRVLCGEWIESMWDCML
Heliotis	FGKNYVD <ins>Y</ins> VDRFPDGDLPRWNFTDFMHSFMIVFRVLCGEWIESMWDCML
Droso	FGKNYHDHKDRFPDGDLPRWNFTDFMHSFMIVFRVLCGEWIESMWDCMY
Musca	FGKNY <ins>ID</ins> HKDRFKD <ins>HE</ins> LPRWNFTDFMHSFMIVFRVLCGEWIESMWDCMY
Myzus	FGKNY <ins>TEK</ins> MYMFKD <ins>HE</ins> LPRWNFTDF <ins>L</ins> HSFMIVFRVLCGEWIESMWDC <ins>LH</ins>
Blattella	FGKNY <ins>YD</ins> NVERFPDGDMPRWNFTDFMHSFMIVFRVLCGEWIESMWDCML
Tutaabsoluta	VGDVSCIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSSLSTPTADNDTN
Plutella	VGDVSCIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSSLSTPTADNETN
Heliotis	VGDVSCIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSSLSTPTADNETN
Droso	VGDVSCIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSSLSAPTADNDTN
Musca	VGDVSCIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSSLSAPTADNDTN
Myzus	VG <ins>EPT</ins> CIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSNLS <ins>V</ins> P <ins>T</ins> ADNETN
Blattella	VGD <ins>W</ins> SCIPFFLATVIGNL <ins>V</ins> VVLNLFLALLLSNFGSSNLSA <ins>P</ins> TADNETN

-S--W--P--T--F--N--L--I--I--S--I--**T**--G--R--T--M--G--A--L--G--
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-N--L--**T**--F--V--L--C--I--I--I--F--I--F--A--V--M--G--M--Q--L--
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-F--G--K--N--Y--V--D--N--V--D--R--F--P--D--G--D--L--P--R--W--
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-I--E--S--M--N--D--C--M--I--V--G--D--V--S--C--I--P--F--F--L--
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-N--F--G--S--S--L--I--T--P--T--A--D--N--D--T--N--K--I--
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862 nt

M918T

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T929I

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|1014F

AAA: Introns

AAA : Exons

AAA : Mutations sites

106 nt



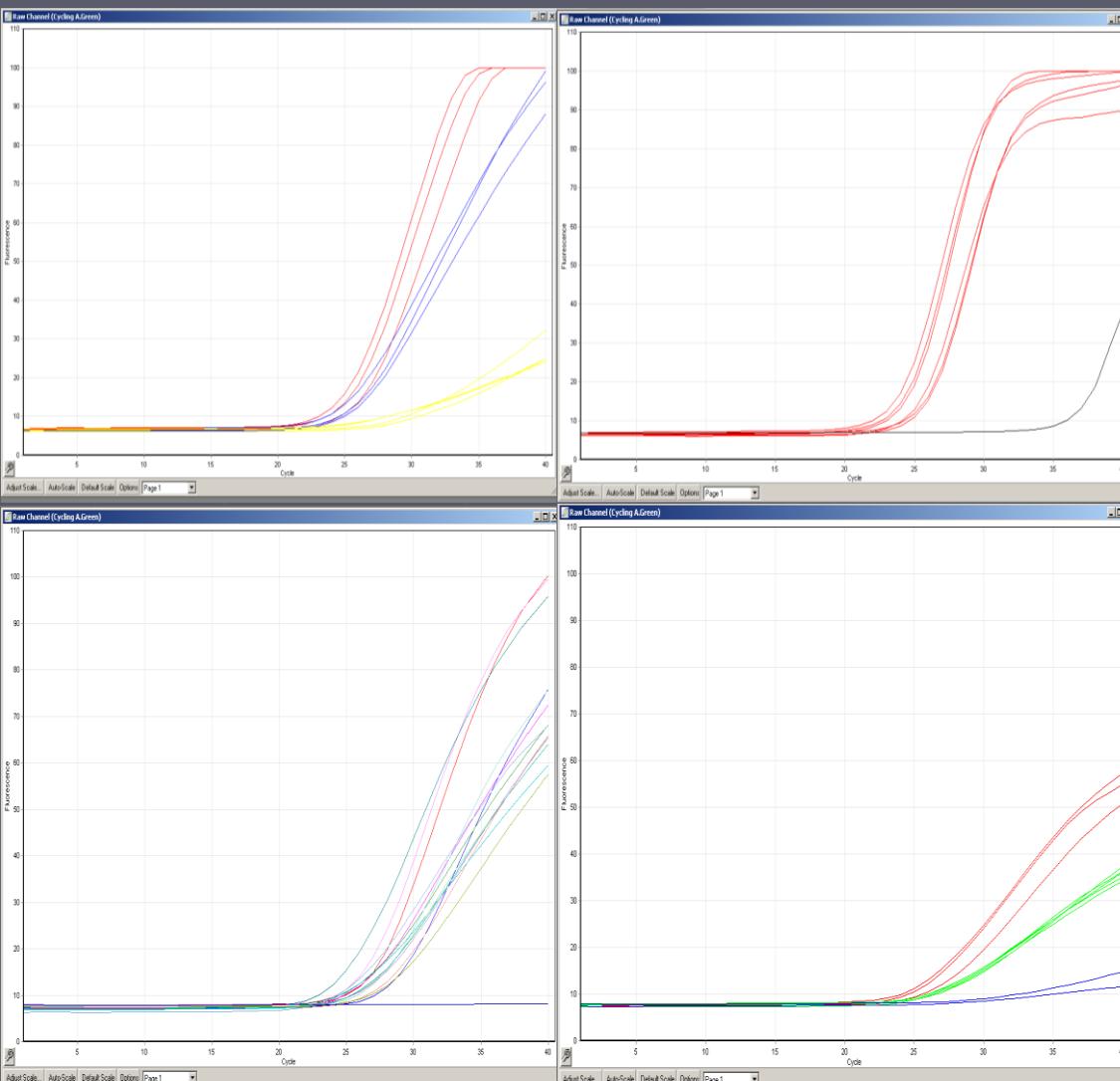
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	2	S/S	R/R		2	S/S	R/R		2	S/S	R/R
	3	R/S	R/S		3	S/S	R/R		3	R/S	R/S
	4	R/S	R/S		4	S/S	R/R		4	S/S	R/R
	5	R/S	R/S		5	S/S	R/R		5	S/S	R/R
	6	R/S	R/S		6	R/S	R/S		6	R/S	R/S
	7	S/S	R/R		7	R/S	R/S		7	S/S	R/R
	8	S/S	R/R		8	R/S	R/S		8	R/S	R/S
	9	R/S	R/S		9	S/S	R/R		9	S/S	R/R
	10	R/R	S/S		10	R/S	R/S		10	R/S	R/S
TA2	1	S/S	R/R	TA4	1	R/S	R/S				
	2	S/R	S/R		2	R/S	R/S				
	3	R/R	S/S		3	R/S	S/S				
	4	R/S	R/S		4	S/S	R/R				
	5	R/R	S/S		5	R/S	R/S				
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	8	R/S	R/S		8	R/S	R/S				
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1824 nt

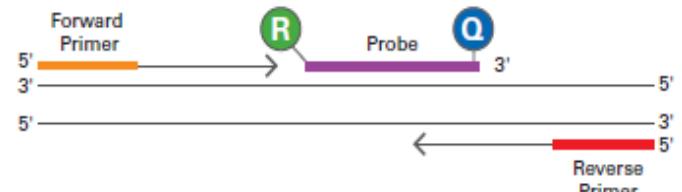
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Consensus	(456)	521	585
GaR3_premix (c)	(1) ---AGGCTTGCATGGTAAAGATAATTGCTATTTCGGAGGTACCCACACAATGTA	ACT	
Ta1R3_premix (c)	(1) ---AGGCTTGCATGGTAAAGATAATTGCTATTTCGGAGGTACCCACACAATGTA	ACT	
Ta2R3_premix (c)	(13) CCATCGCTTGCATGGTAAAGATAATTGCTATTTCGGAGGTACCCACACAATGTA	ACT	
Ta4R3_premix (c)	(1) ----CGCTTGCATGGTAAAGATAATTGCTATTTCGGAGGTACCCACACAATGTA	ACT	
Ta3R3_premix (c)	(1) --TAGGCTTGCATGGTAAAGATAATTGCTATTTCGGAGGTACCCACACAATGTA	ACT	
Torpedo mature protein Consensus	(521) GGATGSCACTGCAGTGGTCAAGAACATCCAGTTCCTCGCGGGACCCAGACGTTGACC		
	(521) AGGCTTGCATGGTAAAGATAATTGCTATTTCGGAGGTACCCACACAATGTA	ACT	
	586	650	
GaR3_premix (c)	(63) TTGTTTGTGAATCATCTGGTGCAGCGTCTGTATCACTTCATTTGTCCTCCATTGTCTAGAAA		
Ta1R3_premix (c)	(63) TTGTTTGTGAATCATCTGGTGCAGCGTCTGTATCACTTCATTTGTCCTCCATTGTCTAGAAA		
Ta2R3_premix (c)	(78) TTGTTTGTGAATCATCTGGTGCAGCGTCTGTATCACTTCATTTGTCCTCCATTGTCTAGAAA		
Ta4R3_premix (c)	(62) TTGTTTGTGAATCATCTGGTGCAGCGTCTGTATCACTTCATTTGTCCTCCATTGTCTAGAAA		
Ta3R3_premix (c)	(64) TTGTTTGTGAATCATCTGGTGCAGCGTCTGTATCACTTCATTTGTCCTCCATTGTCTAGAAA		
Torpedo mature protein Consensus	(586) ATCTCGAGAGTGC CGC GGC GCG TCT GT CGG CA TG C A T T C T C C C G G G G A G G C G A G A		
	(586) TTGTTTGTGAATCATCTGGTGCAGCGTCTGTATCACTTCATTTGTCCTCCATTGTCTAGAAA		
	651	715	
GaR3_premix (c)	(128) TTACCTTTCTCAAGCCATTATGCAGCTGGAGCACGTAACGTACCATGGCTATAAATATCGCGAG		
Ta1R3_premix (c)	(128) TTACCTTTCTCAAGCCATTATGCAGCTGGAGCACGTAACGTACCATGGCTATAAATATCGCGAG		
Ta2R3_premix (c)	(143) TTACCTTTCTCAAGCCATTATGCAGCTGGAGCACGTAACGTACCATGGCTATAAATATCGCGAG		
Ta4R3_premix (c)	(127) TTACCTTTCTCAAGCCATTATGCAGCTGGAGCACGTAACGTACCATGGCTATAAATATCGCGAG		
Ta3R3_premix (c)	(129) TTACCTTTCTCAAGCCATTATGCAGCTGGAGCACGTAACGTACCATGGCTATAAATATCGCGAG		
Torpedo mature protein Consensus	(651) CCTCTTCCGCGGCCATCTTCAGAGCGGCTCGCCAATGCCGTGGCGCTGTCCTGTTG		
	(651) TTACTTTCTCAAGCCATTATGCAGCTGGAGCACGTAACGTACCATGGCTATAAATATCGCGAG		
	716	780	
GaR3_premix (c)	(193) AAGAAAGCATTAAAGGAATTCTTGGCGAAGCTGTACATTGTCCGTACTCAAGAACGAT		
Ta1R3_premix (c)	(193) AAGAAAGCATTAAAGGAATTCTTGGCGAAGCTGTACATTGTCCGTACTCAAGAACGAT		
Ta2R3_premix (c)	(208) AAGAAAGCATTAAAGGAATTCTTGGCGAAGCTGTACATTGTCCGTACTCAAGAACGAT		
Ta4R3_premix (c)	(192) AAGAAAGCATTAAAGGAATTCTTGGCGAAGCTGTACATTGTCCGTACTCAAGAACGAT		
Ta3R3_premix (c)	(194) AAGAAAGCATTAAAGGAATTCTTGGCGAAGCTGTACATTGTCCGTACTCAAGAACGAT		
Torpedo mature protein Consensus	(716) CTGAAGCGCGCAGGGCGGTCAGCTGG--GAAGAA--ACCTCAACTGTAACTC--AACAGC		
	(716) AAGAAAGCATTAAAGGAATTCTTGGCGAAGCTGTACATTGTCCGTACTCAAGAACGAT		
	781	845	
GaR3_premix (c)	(258) GTGGGCCGATGATAAGATGTTACGCCAAAAAACACCTGAAGAACCTTGTAACAATGAATGGGG		
Ta1R3_premix (c)	(258) GTGGGCCGATGATAAGATGTTACGCCAAAAAACACCTGAAGAACCTTGTAACAATGAATGGGG		
Ta2R3_premix (c)	(273) GTGGGCCGATGATAAGATGTTACGCCAAAAAACACCTGAAGAACCTTGTAACAATGAATGGGG		
Ta4R3_premix (c)	(257) GTGGGCCGATGATAAGATGTTACGCCAAAAAACACCTGAAGAACCTTGTAACAATGAATGGGG		
Ta3R3_premix (c)	(259) GTGGGCCGATGATAAGATGTTACGCCAAAAAACACCTGAAGAACCTTGTAACAATGAATGGGG		
Torpedo mature protein Consensus	(775) GACGAAGCTCACTCACTGTCTGGGGAAAAGAGCCTAGGTTGATTGACGTGGAGTGGAA		
	(781) GTGGGCCGATGATAAGATGTTACGCCAAAAAACACCTGAAGAACCTTGTAACAATGAATGGGG		
	846	910	
GaR3_premix (c)	(323) TA-----CATTAGGCATT-TGGAATTTCCT---TTTGTTCCTATTGATGGGTCAATTCTTAG		
Ta1R3_premix (c)	(323) TA-----CATTAGGCATT-TGGAATTTCCT---TTTGTTCCTATTGATGGGTCAATTCTTAG		
Ta2R3_premix (c)	(338) TA-----CATTAGGCATT-TGGAATTTCCT---TTTGTTCCTATTGATGGGTCAATTCTTAG		
Ta4R3_premix (c)	(322) TA-----CATTAGGCATT-TGGAATTTCCT---TTTGTTCCTATTGATGGGTCAATTCTTAG		
Ta3R3_premix (c)	(324) TA-----CATTAGGCATT-TGGAATTTCCT---TTTGTTCCTATTGATGGGTCAATTCTTAG		
Torpedo mature protein Consensus	(840) TGTCTTCCCTTGACAGTATCTCGGTTCTCTCGTCCCGTCATCGATGGGAATTCTTCC		
	(846) TA CATTAGGCATT TGGAATTTCCT TTTGTTCCTATTGATGGGTCAATTCTTAG		
	911	975	

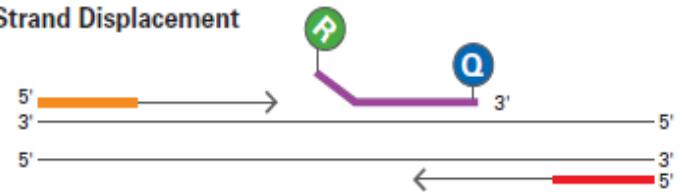
Ala → Ser (position 201):
Aphis gossypii,
Chilo suppressalis,
Plutella xylostella,
Tetranychus urticae,
Bactrocera dorsalis



Polymerization



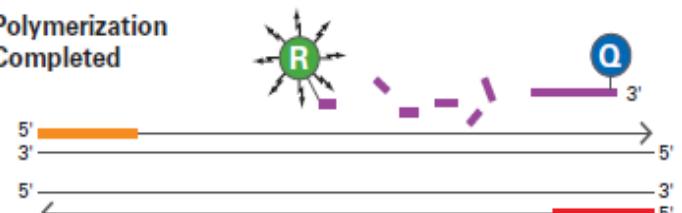
Strand Displacement



Cleavage



Polymerization Completed



Sample ID Nº	Origin	Sampled from
1	Tejina, (La Laguna) Tenerife,. Canary Islands	tomato
2	Guía de Isora, Tenerife, Canary Islands	tomato
3	Granadilla, Tenerife, Canary Islands	tomato
4	Arico,Tenerife, Canary Islands	tomato
5	Teulera, Mallorca, Is Baleares	tomato
6	San Fangos, Mallorca, Is.Baleares	tomato
7	Mostaganem, Algeria	Tomato under-protection
8	Mostaganem, Algeria	Tomato under-protection
9	Turín Italy	Unknown
10	La Tola, Pichincha, Ecuador	tomato
11	Tudela, Navarra, SPAIN	tomato
12	Barrancas,Santa Fé, Argentina	Tomato under-protection
13	LaPrimavera, Mendoza, Argentina	tomato
14	La Plata, Bs.As. Argentina	Tomato under-protection
15	Cagliari.S.Margherita di Pula/ ITALY	Tomato under-protection
16	La Palma, Canary Islands	Unknown
17	Sicilia ITALY	Aubergine
18	Ramonete/Lorca, MU, SPAIN	tomato
19	(Curicó costa), CHILE	tomato
20	Antioquía/Rionegro/ Colombia	tomato
21	Heraklion/Creta- Greece	wild plants
22	Mazarrón, Murcia, SPAIN	tomato
23	Canelones; URUGUAY	tomato
24	Chulacanas, Piura. Perú	tomato
25	Valencia, SPAIN	Unknown
26	Maresme, Cataluña, SPAIN	tomato
27	Silveira. Concello Torres Vedres, Portugal	tomato
28	Sele valley.Salerno.Campania. ITALY	tomato
29a	beit hashita, israel valley (1)	Unknown
29b	ein hmifraz, western galilee (2)	Unknown

Conclusions:

- All the five strains showed differences in susceptibility to the compounds used with LC50 far higher than the recommended field rates.
- A 420 b fragment of sodium channel (domains IIS5/IIS6) and 1824 b fragment of the acetylcholinesterase of *Tuta absoluta* were sequenced for the first time.
- All the five strains have the kdr (L1014F) mutation in combination with either super kdr(M918T) or T929I mutation.
- A rare novel mutation L925M was detected in 2 individuals from one strain.
- All the strains showed the Ala to Ser mutation at the position 201 of acetylcholinesterase.
- A TaqMan was designed to screen field collected samples of diverse geographic origin and to examine the frequency and distribution of the reported mutations.

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Thank You